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APPLICATION NO. 09/7463881 (Serial No. 7 CFR 15)

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INTERNATIONAL APPLICATION NO.	INTERNATIONAL FILING DATE	PRIORITY DATE CLAIMED
PCT/GB98/02316	31 July 1998	31 July 1997
TITLE OF INVENTION: METHOD AND APPARATUS FOR DETERMINING MOLECULAR CRYSTAL STRUCTURES		
APPLICANT(S) FOR DO/EO/US: DAVID, William I. And SHANKLAND, Kenneth		
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:		
<p>1. <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. 371.</p> <p>2. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371.</p> <p>3. <input checked="" type="checkbox"/> This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371 (b) and PCT Articles 22 and 39(1).</p> <p>4. <input checked="" type="checkbox"/> A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.</p> <p>5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371(c)(2)) a. <input checked="" type="checkbox"/> is transmitted herewith (required only if not transmitted by the International Bureau). b. <input type="checkbox"/> has been transmitted by the International Bureau. c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US).</p> <p>6. <input type="checkbox"/> A translation of the International Application into English (35 U.S.C. 371(c)(2)).</p> <p>7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)) a. <input type="checkbox"/> are transmitted herewith (required only if not transmitted by the International Bureau). b. <input type="checkbox"/> have been transmitted by the International Bureau. c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired. d. <input checked="" type="checkbox"/> have not been made and will not be made.</p> <p>8. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 34. a. <input type="checkbox"/> are transmitted herewith (required only if not transmitted by the International Bureau). b. <input type="checkbox"/> have been transmitted by the International Bureau. c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired. d. <input checked="" type="checkbox"/> have not been made and will not be made.</p> <p>9. <input type="checkbox"/> A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).</p> <p>10. <input checked="" type="checkbox"/> An unsigned oath or declaration from the inventor. (35 U.S.C. 371 (c)(4)).</p> <p>11. <input type="checkbox"/> A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).</p>		
Items 12. to 16. below concern other document(s) or information included:		
<p>12. <input checked="" type="checkbox"/> An information Disclosure Statement under 37 CFR 1.97 and 1.98.</p> <p>13. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.</p> <p>14. <input checked="" type="checkbox"/> A FIRST preliminary amendment. <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment.</p> <p>15. <input type="checkbox"/> A substitute specification.</p> <p>16. <input type="checkbox"/> A change of power of attorney and/or address letter.</p> <p>17. <input checked="" type="checkbox"/> Other items or information: Certificate of Express Mailing Under 37 CFR 1.10</p>		

PATENT
Attorney Docket No.: 9267-8

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE
ACTING AS THE DESIGNATED/ELECTED OFFICE

In re: Patent application of : Group Art Unit:
William I. David and Kenneth Shankland : Not Yet Assigned
U.S. Serial No.: Not Yet Assigned : Examiner:
International Application No.: : Not Yet Assigned
PCT/GB98/02316 : International Filing Date:
Filed: Concurrently Herewith : 31 July 1998
For: **Method and Apparatus For Determining**
Molecular Crystal Structures :

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
BOX PCT
Washington, D.C. 20231

Attention: US/DO/EO

Dear Sir:

Prior to examination of this application and before calculation of the filing fee,
please amend the application, without prejudice, in accordance with the following.

Charge any fee or credit any overage associated with this preliminary amendment
or the application filing to Deposit Account No. 19-1135.

* * *
CERTIFICATE OF MAILING
UNDER 37 C.F.R. 1.10

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Jayne Weller
Signature of person mailing paper

Jayne Weller
Type or print name of person

Please amend the application, without prejudice, in accordance with the following.

In the Claims:

Re-write the claims as follows:

4. (Amended) The method as claimed in [either of claims 2 or 3] claim 2, wherein the total data in the reduced representation is reduced by a factor substantially equal to the number of data points in the original powder diffraction data divided by the number Bragg reflections in the measured data range.

5. (Amended) The method as claimed in [either of claims 3 or 4] claim 2, wherein the fitness χ^2 of each of the trial structures is determined using the following function:

$$\chi^2 = \sum_h \sum_k \{ (I_{h,k} - c |F_{h,k}|^2) (V^{-1})_{hk} (I_{h,k} - c |F_{h,k}|^2) \}$$

where:

$I_{h,k}$ = extracted intensity

$V_{h,k}$ = covariance matrix

c = a scale factor

$F_{h,k}$ = calculated structure factor from trial structure

6. (Amended) The method as claimed in [any one of the preceding claims] claim 1, wherein the set of variables consists of three coordinates representative of the location of the molecule within the unit cell and three independent coordinates representative of the orientation of the molecule within the unit cell.

8. (Amended) The method as claimed in [any one of the preceding claims] claim 1 including the step of determining the unit cell and space group for the molecule under examination.

9. (Amended) The method as claimed in [any one of the preceding claims] claim 1 including the step of determining the set of internal coordinates.

10. (Amended) The method as claimed in [any one of the preceding claims] claim 1, further including the step of monitoring the number of iterations in which new trial structures are generated and halting the method and outputting the trial crystal structure with the best calculated fitness after completion of a predetermined number of iterations.

11. (Amended) The method as claimed in [any one of the preceding claims] claim 1, wherein the selection of survivors and the alteration of the values of the variables is based on a simulated annealing procedure.

15. (Amended) The method as claimed in [either of claims 13 or 14] claim 13, wherein the total data in the reduced representation is reduced by a factor substantially equal to the number of data points in the original powder diffraction data divided by the number Bragg reflections in the measured data range.

16. (Amended) Apparatus as claimed in [either of claims 14 or 15] claim 14, wherein the fitness analyser determines the fitness χ^2 of each of the trial structures using the following function:

$$\chi^2 = \sum_h \sum_k \{ (I_h - c |F_h|^2) (V^{-1})_{hk} (I_k - c |F_k|^2) \}$$

where:

$I_{h,k}$ = extracted intensity from the structure factor analyser

$V_{h,k}$ = covariance matrix from the structure factor analyser

c = a scale factor

$F_{h,k}$ = calculated structure factor from trial structure

17. (Amended) Apparatus as claimed in [any one of claims 12 to 16] claim 12, wherein controller determines a set of variables consists of three coordinates representative of the location of the molecule within the unit cell and three independent coordinates representative of the orientation of the molecule within the unit.

18. (Amended) Apparatus as claimed in [any one of claims 12 to 17] claim 12, wherein the structure factor analyser additionally determines the unit cell and space group for the molecule under examination.

19. (Amended) Apparatus as claimed in [any one of claims 12 to 18] claim 12 including a coordinate generator for determining the set of internal coordinates.

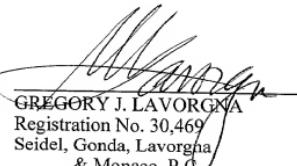
20. (Amended) Apparatus as claimed in [any one of claims 12 to 19] claim 12, further including a counter for monitoring the number of iterations of new trial structures.

REMARKS

The above amendments are intended to remove multiple dependency within the claims prior to entry into the U.S. national phase. Applicant hereby requests entry of the above amendments prior to calculation of the filing fee and examination.

Respectfully submitted,

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Applicant or Patentee: William Ian David and Kenneth Shankland

Serial or Patent No.: Not Yet Assigned

International Application No.: PCT/GB98/02316

Submitted To DO/EO/US: January 31, 2000

For: Method and Apparatus For determining Molecular Crystal Structures

**VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS
(37 CFR 1.9(f) AND 1.27(d)) - NONPROFIT ORGANIZATION**

I hereby declare that I am an official empowered to act on behalf of the nonprofit organization identified below:

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ADDRESS OF ORGANIZATION Chilton, Didcot, Oxfordshire OX11 0QX Great Britain

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(NAME OF STATE _____)
(CITATION OF STATUTE _____)

I hereby declare that the nonprofit organization identified above qualifies as a nonprofit organization as defined in 37 CFR 1.9(e) for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United State Code with regard to the above-identified invention by inventor(s) identified above

the Specification filed herewith.

Application Serial No. Not Yet Assigned submitted to DO/EO/US 31 January 2000
International Application No. PCT/GB98/02316 filed 31 July 1998.

Patent No. _____, issued _____.

I hereby declare that rights under contract or law have been conveyed to and remain with the nonprofit organization with regard to the above identified invention.

If the rights held by the nonprofit organization are not exclusive, each individual concern or organization having rights to the invention is listed below* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR 1.9(d) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

*NOTE: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities.
(37 CFR 1.27)

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING LINDA BAINES

TITLE IN ORGANIZATION: Head, Sales Contracts

ADDRESS OF ORGANIZATION CCLRC Rutherford Appleton

Laboratory

SIGNATURE Linda Baines

DATE 8-02-00

METHOD AND APPARATUS FOR DETERMINING MOLECULAR
CRYSTAL STRUCTURES

The present invention provides an improved method and apparatus
5 for determining molecular crystal structures from powder diffraction data. In particular, the present invention enables molecular crystal structures to be identified using only powder diffraction data in a manner that is considerably faster than is currently the case. Furthermore, with the
10 present invention the molecular crystal structure of large organic molecules, such as pharmaceutical compounds, can be determined using data from powder diffraction analysis.

Information on the molecular crystal structure of a molecule is usually obtained through irradiation of a single crystal of the molecule with neutrons or X-rays. Subsequent analysis of the resultant diffraction
15 pattern, which consists of a series of angularly spaced intensity peaks with each peak representing an individual Bragg reflection, provides information on the structure. Whilst this single crystal diffraction technique is an effective technique for determining the crystal structure of a molecule, it can often prove difficult to grow the single crystals necessary for the
20 analysis. Moreover, where the molecule can crystallise in more than one polymorphic form, it is sometimes the case that it can prove very difficult to grow a single crystal of a particular polymorph.

To address these problems, a powder diffraction analysis was developed in which a crystalline powder of the material under analysis is
25 irradiated instead of a single crystal. Analysis of the resultant diffraction pattern is hampered by the fact that the diffraction pattern may include Bragg reflections that partially or fully overlap one another, making it difficult for individual reflections to be identified, and their associated intensities quantified. An example of experimental data from irradiation of
30 a powder sample of the drug substance cimetidine in the form of a graph representing intensity of the Bragg reflections with respect to angular position is shown in Figure 1. Currently, in order to identify molecular

crystal structures diffraction patterns of this type are used in a point-to-point comparison with diffraction data calculated from a postulated model of the crystal structure. If there is good agreement between the measured and calculated diffraction data, it may be assumed that the postulated

5 structure is close to the true crystal structure of the molecule. In general, good agreement is only obtained when there is significant prior knowledge of the true crystal structure of the molecule, as there is an infinite number of crystal structures that may be postulated and compared to the experimental data. Moreover, such analysis is very slow and time

10 consuming because of the extremely large body of point-to-point data that must be compared for each postulated crystal model in turn.

The present invention seeks to address the problems discussed above with respect to existing diffraction analysis techniques and in particular to provide a method and apparatus that is considerably faster

15 than conventional techniques. The present invention further seeks to provide a method and apparatus for determining molecular crystal structures which employs data obtained from the irradiation of crystalline powders and permits analysis without the need for prior knowledge of any approximate crystal structure.

20 The present invention provides a method for determining molecular crystal structures from powder diffraction data comprising the steps of: generating a reduced representation of the powder diffraction pattern in dependence on a predetermined unit cell and space group of the molecule under examination in which the total quantity of diffraction data is

25 significantly reduced whilst maintaining the characteristics of the diffraction data that are representative of the crystal structure under examination; determining a set of variables for describing trial molecular structures, derived from predetermined internal co-ordinates and said space group; assigning values to said variables thereby creating a population of trial

30 structures each defined by a unique set of values for said variables; calculating a fitness for each trial structure with respect to the reduced representation of the powder diffraction pattern; determining whether any

one of the calculated fitnesses is less than or equal to a predetermined threshold; where none of the calculated fitnesses is less than or equal to the threshold value, selecting at least one survivor from the population of trial structures, altering the values of the variables of at least one of the

5 survivors in accordance with one or more predetermined rules, calculating the fitnesses of the new trial structures; and repeating the steps of selecting survivors, altering the values of the variables and calculating the fitnesses of the new trial structures until at least one of the calculated fitnesses is less than or equal to the threshold value, and where at least

10 one of the calculated fitnesses is less than or equal to the threshold, outputting at least one trial molecular crystal structure represented by the successful sets of values.

In a further aspect the present invention provides apparatus for determining molecular crystal structures comprising a structure factor analyser for generating from experimental powder diffraction data for the molecule under examination a reduced representation of the powder diffraction pattern based on a predetermined unit cell and space group in which the total quantity of diffraction data is significantly reduced whilst the characteristics of the diffraction data representative of the crystal structure

15 under examination are maintained; a controller for determining a set of variables for describing trial molecular structures, derived from predetermined internal co-ordinates and said space group; a searching processor for creating a population of trial structures each defined by a unique set of values for said variables said searching processor including a

20 fitness analyser for calculating a fitness for each trial structure with respect to the reduced representation of the powder diffraction pattern, a thresholding device for determining whether any one of the calculated fitnesses is less than or equal to a predetermined threshold, a survivor selector for selecting at least one survivor from the population of trial

25 structures, a variable adjustment device for altering the values of the variables of at least one of the survivors and output means for outputting the one or more trial molecular crystal structures having calculated

30

fitnesses less than or equal to the threshold value.

The reduced representation preferably consists of identification of each reflection in the powder diffraction data along with associated weighting factors and ideally is in the form of a structure factor intensity

5 listing and associated covariance matrix. Reference herein to reflections is intended as reference to individual Bragg reflections or peaks in the diffraction data resulting from reflections of the incident radiation from the structure of the molecule.

10 Moreover, preferably, the fitness χ^2 of each of the trial structures is determined using the following function:

$$\chi^2 = \sum_h \sum_k \{ (I_h - c|F_h|^2) (V^{-1})_{hk} (I_k - c|F_k|^2) \}$$

where:

15 $I_{h,k}$ = extracted intensity from the structure factor analyser
 V_{hk} = covariance matrix from the structure factor analyser
 c = a scale factor
 $F_{h,k}$ = calculated structure factor from trial structure

20 The plurality of co-ordinates preferably consist of three co-ordinates representative of the location of the molecule within the unit cell, three co-ordinates representative of the orientation of the molecule within the unit cell and one or more co-ordinates representative of respective torsion angles.

25 In a preferred embodiment the structure factor analyser additionally automatically determines the optimal unit cell and space group for the molecule under examination instead of the unit cell and space group being predetermined manually and input into the structure factor analyser. More, preferably a co-ordinate generator is provided for automatically determining 30 the set of internal co-ordinates instead of the set of internal co-ordinates being predetermined manually and input into the controller.

The search for a three dimensional structure of a molecule which

would produce a powder diffraction pattern nearly identical to available experimental results is greatly simplified with the present invention and considerably speeded up by a factor of typically up to 1000 or more. This is achieved by the reduction of trial molecular crystal structures to a unique

5 set of variables that does not explicitly include the co-ordinates of individual atoms in the molecule but instead includes a single set of co-ordinates for the location and orientation of the entire molecule. Reduction of the experimental powder diffraction data to data representative of each reflection in combination with a weighting factor also assists in speeding up

10 the analysis of trial crystal structures in which the fitness of each trial structure with respect to the reduced experimental data is determined using structure factor analysis.

The present invention relies on the fact that at its most basic, a molecular crystal structure can be represented by a set of internal co-ordinates describing the molecule under investigation together with co-ordinates describing the location and orientation of the molecule within a unit cell of which only some but not all need be variable. The reduction of the molecular crystal structure to such a set of variables enables analysis of the trial structures to be performed much more quickly than an analysis

15 20 performed using the conventional method of describing the crystal structure in terms of the fractional or Cartesian co-ordinates of every atom in the asymmetric unit of the structure. Such conventional representations are considered to be unworkable in a model building sense because of the computing power necessary to position individual atoms independently of each other.

The representation of the trial structures used in the invention along with the novel fitness function means that analyses can be performed in seconds or minutes on the current generation of conventional personal computers or workstations. Moreover, the representation is versatile as it

25 30 allows the invention to work with flexible molecules as well as multiple fragments.

An embodiment of the present invention will now be described by

way of example with reference to the accompanying drawings, in which:

Figure 1 is a graph of experimental data from x-ray powder diffraction analysis of cimetidine showing 315 reflections, using an irradiation wavelength of 1.5285 \AA and a data range for 2 θ of 8 $^{\circ}$ -56 $^{\circ}$;

5 Figure 2 is a schematic representation of the 2D molecular structure of cimetidine;

Figure 3 is a flow diagram of the method steps for determining a molecular structure in accordance with the present invention;

Figure 4 is a diagram of the crystal structure of cimetidine;

10 Figures 5a, 5b, 5c and 5d are diagrams showing the progressive development of a trial crystal structure for cimetidine, employing the method and apparatus in accordance with the present invention, overlying the diagram of Figure 4;

15 Figure 6 is a graph showing the fitness of a trial crystal structure for cimetidine with respect to generations, employing the method and apparatus in accordance with the present invention;

Figures 7a, 7b and 7c show the molecular structure of dopamine deuterobromide, a graph of the development of a trial structure for the crystal and a diagram of the solution respectively, employing the method and apparatus of the present invention;

20 Figure 8 is a graph of experimental data from x-ray powder diffraction analysis of capsaicin using an irradiation wavelength of 0.6528 \AA and a data range for 2 θ of 2.7 $^{\circ}$ -22.5 $^{\circ}$;

25 Figure 9 is a tabulation of part of the reduced representation of the powder diffraction data of Figure 8 generated in accordance with the method of the present invention; and

30 Figure 10 is a diagram comparing the crystal structure of capsaicin obtained from single crystal diffraction data with the crystal structure obtained using powder diffraction data alone in a method in accordance with the present invention.

The present invention will be described with reference to an experimental determination of the crystal structure of the molecule

cimetidine, a histamine H₂ antagonist used in the treatment of stomach ulcers, for which a full single crystal structure (monoclinic Form A) determination has already been performed. Figure 2 shows the 2D chemical formula of the cimetidine molecule, whilst the known arrangement 5 of the cimetidine molecules within the unit cell of the crystal structure is shown in Figure 4.

To determine the molecular crystal structure of cimetidine employing the method and apparatus of the present invention with reference to Figure 3, initially a conventional powder diffraction pattern (10) is obtained from a 10 crystalline powder sample of cimetidine. The resultant diffraction pattern is shown in Figure 1. The experimental diffraction data (10) is input into a cell dimension analyser (12). The cell dimension analyser (12) uses conventional techniques to assess the diffraction pattern in order to determine the unit cell dimensions of the crystal structure. The generation 15 of the unit cell dimensions may alternatively be performed manually, however, it is preferred that the unit cell dimensions be generated automatically using the crystal modelling apparatus. The diffraction pattern is also input to a structure factor analyser (14) that also receives the unit cell dimensions determined by the analyser (12). The structure factor 20 analyser (14) analyses the experimental diffraction pattern using the lowest symmetry space group consistent with the crystal class determined by the cell dimension analyser (12), reducing the data to a first structure factor intensity listing and an associated covariance matrix. From this listing, the true space group (16) of the crystal structure is determined and used by 25 the structure factor analyser (14) to generate a second structure factor intensity listing and associated covariance matrix (18). By generating this second structure factor intensity listing and associated covariance matrix (18), the total quantity of the original experimental diffraction data is significantly reduced in amount without loss of those characteristics of the 30 data representative of the crystal structure under examination. The original data can be reduced by a factor typically equal to the number of data points in the original powder diffraction data divided by the number Bragg

reflections in the measured data range. Thus, the experimental diffraction data is not presented for analysis as a point-by-point profile, but rather in a reduced data form enabling the data to be analysed using a fitness function described in greater detail below. In performing the reduction of 5 the original data the individual Bragg reflections (peaks) are identified and each reflection is allocated a value representative of its intensity along with one or more factors that describe the extent to which the reflection overlaps with one or more adjacent reflections.

As an example of the extent to which the powder diffraction data is 10 reduced, typically powder diffraction data will contain between 2000 and 5000 individual data points with the reduced representation of the diffraction data the total number of data points is reduced to between 100 and 400. Hence, the original data is reduced by a factor of between 10 and 30 whilst still retaining the characteristics of the original data. 15 Furthermore, the speed of analysis of each individual data point is increased over conventional techniques because with the original powder diffraction data the analysis of each data point includes not only the structure factor but also a factor representative of the shape of the peak. With the reduced data the analysis of each data point is reduced to only 20 the structure factor.

Where the selected space group based on the unit cell dimensions does not prove to be the correct space group, the method described above is repeated using different space groups until the correct space group is identified.

25 Using the known 2D chemical formula for cimetidine (20), a co-ordinate generator (22) determines a set of internal co-ordinates (24) which completely describe the three dimensional structure of the molecule. The internal co-ordinates (24) include known data using tabulated bond lengths, bond angles and rigid torsion angles, where necessary, along with 30 identification of unknown variables such as flexible torsion angles. When postulated values for the unknown variables are added, sufficient information is present in the internal co-ordinates to define the

conformation of an isolated theoretical cimetidine molecule.

Preferably, the only unknown factors and so the only variables to be found in the internal co-ordinates are the values of the variable torsion angles (represented by variables $\tau_1, \tau_2, \tau_3, \tau_4, \tau_5 \dots$). It is not essential for 5 the bond lengths and bond angles to be held fixed and where appropriate these factors too may be varied in determining the crystal structure of the molecule. It has been found though that variation of the bond lengths and bond angles within chemically sensible bounds has a much smaller effect on the calculated diffraction data than variation of the flexible torsion 10 angles within the structure. Thus for most purposes, acceptable results can be achieved with these factors held fixed.

Generation of the set of internal co-ordinates may alternatively be performed manually in which case the manually generated set of internal co-ordinates is input into the crystal modelling apparatus.

15 The output (24) of the co-ordinate generator (22) is supplied to a controller (26) that is also connected to the space group output (16) of the structure factor analyser (14). The controller (26) also includes an input (28) to enable manual setting of selected operational parameters such as the number of trial structures to be analysed in each generation, i.e. the 20 population size. The controller (26) uses the internal co-ordinates and the space group to determine additional variables representing the location and orientation of a molecular structure in the unit cell. Preferably, the location of the molecular structure within the unit cell is defined using a single reference point in fractional co-ordinate space represented by 25 external co-ordinates or variables (x, y, z). The orientation of the molecule at that point may be described using Euler angles (α, β, γ). Alternatively, the orientation of the molecule may be described using a quaternion, q .

In this way the molecular crystal structure is reduced to a set of variables consisting of internal and external co-ordinates:

30 $\{x, y, z, \alpha, \beta, \gamma, \tau_1, \tau_2, \tau_3, \tau_4, \tau_5 \dots\}$ or $\{x, y, z, q, \tau_1, \tau_2, \tau_3, \tau_4, \tau_5 \dots\}$.

These variables are suitable for iterative mathematical processing and are more amenable to search procedures than the full complement of

individual atomic co-ordinates used in conventional techniques.

The output (30) from the controller (26) is then supplied to an iterative searching processor (32). The output (30) consists of the set of variables determined by the controller (26); the complete internal co-
5 ordinates produced by the co-ordinate generator (22); operating parameters such as the selected size of the population to be employed in the searching procedure; any rules restricting or controlling the values which can be allocated to each of the variables; and any rules controlling the selection of survivors, the breeding and the mutation of survivors,
10 described in greater detail below.

In the method shown in Figure 3, the iterative searching processor (32) employs a genetic algorithm to determine the correct molecular crystal structure. The above mentioned set of variables $\{x, y, z, \alpha, \beta, \gamma, \tau_1, \tau_2, \tau_3, \tau_4, \tau_5 \dots\}$ or $\{x, y, z, q, \tau_1, \tau_2, \tau_3, \tau_4, \tau_5 \dots\}$ are thus equated to chromosomes,
15 with each individual variable equating to a gene. The genetic algorithm establishes certain protocols based on the concept of 'survival of the fittest', with respect to the selection of survivors, the breeding and the mutation of survivors.

Firstly, within the searching processor (32) an initial population of chromosomes is created (33) by assigning random numbers to each of the genes of each of the chromosomes. The allowable random numbers for any particular gene may be restricted in accordance with rules input from the controller (26). The selected size of this initial population depends somewhat upon the complexity of the structure under investigation, with
20 larger population sizes typically being required for problems involving more variables. In the case of cimetidine, where seven torsion angles were allowed to vary, resulting in thirteen degrees of freedom, a population size of 150 was chosen. The fractional co-ordinates (x, y, z) and Euler angles are randomly set as real numbers normally bounded by the Euclidian
25 normalisers of the space group. The variable torsion angles (τ) are typically randomly set as real numbers in the range 0°-360°.

Using the internal co-ordinates a three dimensional structure of the

trial molecule is constructed (35) for each parent in Cartesian space, and then in fractional space with respect to the crystal unit cell. Diffraction data is then determined (37) for each of the trial molecular structures and a fitness value, χ^2 , is calculated (39) for each trial structure with respect to 5 the structure factor intensity listing and covariance matrix. The preferred fitness function employed is as follows:

$$\chi^2 = \sum_h \sum_k \{ (I_h - c|F_h|^2) (V^{-1})_{hk} (I_k - c|F_k|^2) \}$$

10 where:

$I_{h,k}$ = extracted intensity from the structure factor analyser

V_{hk} = covariance matrix from the structure factor analyser

c = a scale factor

$F_{h,k}$ = calculated structure factor from trial structure

15 In determining the fitness value, χ^2 , the optimal value of the scale factor c must be determined. This is preferably done by performing a conventional linear least squares analysis to determine the optimal scaling factor between the calculated structure factors and the reduced data. In addition to optimising with respect to the scale factor, the fitness value, χ^2 , 20 may also be optimised with respect to quantities such as an overall temperature factor or a preferred orientation parameter in which case a conventional non-linear least squares analysis is preferably performed.

The fitness values for each of the chromosomes is compared to a predetermined threshold value (41) so that in the event any one of the 25 chromosomes is less than or equal to the threshold value a solution for the molecular structure is output (43). In the event that the fitness values of all of the chromosomes exceed the threshold value, the chromosomes are then supplied (45) to a survivor selector (47). At the same time a counter (49) is increased by one so that a record of the number of generations 30 created is maintained.

Using the fitness values obtained for each of the chromosomes, the survivor selector (47) employs a proportional selection scheme, in which

the chances of a chromosome surviving are proportional to its fitness, to select a number of survivors. Other criteria for selecting survivors may alternatively be used. For example, a tournament selection may be employed in which case two chromosomes are selected at random and

5 compared with one another, with the fittest surviving. In particular the Boltzmann tournament may be used as it introduces an element of simulated annealing to the selection process. In addition, the selection may be elitist with the best member of the population in terms of fitness always surviving to enter the next generation.

10 Additional fitness functions may also be employed instead of, or in combination with the aforementioned fitness function, to further enhance the analysis of the trial structures. For example, simultaneous fitting of both X-ray and neutron diffraction data; use of a molecular packing function; use of an isolated molecule Lennard-Jones type calculation; use

15 of a rotation / translation function; and use of phase information derived from direct / Patterson methods.

Although the above method is described in terms of the entire population being subject to a common selection, the population may be divided into sub-populations in which each sub-population evolves

20 independently of the other sub-populations albeit that migration from one sub-population to another can be enabled.

The surviving chromosomes are then used to create offspring (51) by allowing the chromosomes to 'breed'. For example, individual genes from different chromosome survivors may be mixed and/or one or more of

25 the genes in a chromosome survivor or its offspring may be mutated by random selection of a new value for the gene. Often, the population size is kept constant throughout this breeding process. The three dimensional structure of each of the offspring is then determined (35), as before, and theoretical diffraction data calculated (37).

30 The fitness (χ^2) is then evaluated (39) for each of the offspring and the fitness results compared (41) to the predetermined threshold value to determine whether a likely crystal structure for the molecule has been

identified. If one of the offspring chromosomes has a fitness value which is less than or equal to the threshold value, or if a predetermined maximum number of generations has been reached, then the search procedure is stopped (43). On the other hand, if the fitness functions of the 5 chromosome offspring all exceed the threshold value and the counted number of generations is less than the maximum allowed number, then the offspring are returned for the selection of survivors (47) and for the creation of new offspring (51).

Additional rules may also be employed where appropriate to 10 constrain the allowable values for the variables. These rules are determined by the controller (26) that may utilise data on crystal fragments stored in a memory (53). For example, the controller (26) may search through stored crystallographic databases of known crystal structure fragments related to the molecule to provide prior information about torsion 15 angle values likely to be adopted by the structure. Such information can then be implemented either as hard limits on the allowable values the torsion angles may adopt, or as probability distributions for the torsion angles. Furthermore, fragments of the molecule may be located using Patterson methods or direct methods. For example, the location of a 20 heavy atom may be used to anchor a molecule during the analysis by the searching processor (32). This effectively reduces the dimensionality of the problem by three as the fractional reference co-ordinates are then known.

Operation of the processor (32) in the search for the correct 3D 25 molecular crystal structure is thus an iterative procedure with the average fitness for each generation gradually tending towards the global minimum in fitness function space. In Figure 5a, a trial cimetidine crystal structure, corresponding to a chromosome in the first generation initialised at random by the processor (32), is shown overlying the true crystal structure first 30 shown in Figure 4. Figure 5b then shows one of the early offspring determined by the processor having a fitness value of $\chi^2=980$, again overlying the true crystal structure of cimetidine. In Figure 5c, a later

offspring having a fitness value of $\chi^2=430$ is shown and the improvement in correspondence between the trial crystal structure and the actual crystal structure is immediately evident. At this point, the crystal structure could be refined using a conventional constrained Rietveld refinement. Hence,

5 the processor (32) may be arranged so that the threshold value for the fitness function is set at around 450. This would result in the search procedure being stopped once the trial structure shown in Figure 5c had been generated, thereby enabling alternative methods to be used to refine the fine details of the trial structure. The advantage of stopping the search

10 procedure at this point is that, usually, conventional methods will be able to refine the fine details of the structure more efficiently than the presently described method and apparatus.

Continuing with the present method, in Figure 5d an offspring having a fitness value of $\chi^2=110$ is shown at which point the detail of the trial

15 structure is easily refinable. Figure 6 is a graph of trial results for cimetidine using the method described above showing the fitness value of offspring with respect to the number of generations for both average fitness and the best fitness. As can be seen, a refinable structure is obtained within a few hundred generations, and an easily refinable structure is

20 obtained around 3000 generations. This latter structure corresponds to an elapsed time of approximate 40 minutes, with the processor running on a single 175MHz R10000 Silicon Graphics™ workstation.

As further examples for the speed of this method, easily refinable structures for pyrene were determined in around 33 seconds, around 15

25 seconds for chlorothiazide and 36 minutes for Ibuprofen, with all calculations being performed on a single 200MHz Pentium Pro™ personal computer.

The above method and apparatus may also be used with molecular structures consisting of more than one fragment. As shown in Figures 7a,

30 7b and 7c an easily refinable structure solution for dopamine deuterobromide using neutron powder diffraction data was achieved in around only 4000 generations. This structure involves not only a dopamine

cation, but also a separate bromide anion. Using the present method and apparatus the location, orientation and conformation of the cation, and the location of the anion can be determined simultaneously.

Whilst in the examples given above the individual genes are real numbers, they could equally be represented by binary strings or integer approximations with appropriate scaling factors. Also, in the example given above the experimental diffraction data is reduced to a structure factor listing and associated covariance matrix, it will be apparent that alternative ways of reducing the total quantity of diffraction data may be employed which, although providing less faithful representations of the diffraction data, nevertheless preserve sufficient characteristics of the original diffraction data to enable a successful structure determination to be performed. For example the data relating to the correlation of reflections may be omitted from the reduced representation.

In the above example a genetic algorithm searching processor is employed to perform an iterative selection of candidate molecular crystal structures. Alternative iterative analysis processes such as simulated annealing, evolutionary strategies and neural network analysis may be used instead of the genetic algorithm. For example, using a simulated annealing process, only a single member is normally utilised and the same variables that were treated as genes by the genetic algorithm are individually adjusted by a small perturbation of their current values. If the function value (χ^2 as defined previously) is better than before, then the new values of the variables are retained. If the function value is worse, then the new values of the variables are not automatically rejected. Instead the new values may be retained if allowed by the temperature dependent Boltzmann selection protocol. In this way, 'uphill' (in terms of χ^2) adjustment of the variables is permissible, helping the algorithm to escape from local minima. The initial choice of the temperature is usually high to allow large 'uphill' moves if necessary, but the temperature is usually lowered in some predetermined fashion during the iterative process. One such way is a temperature reduction that cools more slowly if the χ^2

fluctuations are large.

An example employing the simulated annealing process to the determination of the crystal structure of capsaicin is shown in Figures 8 to 10. Experimental powder diffraction data for capsaicin is shown in Figure 8 with the first page of a tabulation of the reduced representation of the data produced using the above mentioned method shown in Figure 9. In Figure 10 the crystal structure obtained from powder diffraction data alone is overlaid upon the crystal structure obtained by the conventional single crystal diffraction route is shown. The experimental powder diffraction data 10 consists of 9901 data points whilst the reduced representation of this data as exemplified in Figure 9 contains only 379 data points, in total.

Figure 9 shows the first page of the tabulated reduced representation consisting of twelve columns with columns A to C identifying the Miller indices of the Bragg reflections; column D identifying the Bragg 15 intensity; column E an esd for the Bragg intensity; column F provides a numerical count of the separable reflections whereby identical numbers in consecutive rows indicates where reflections are so close together they cannot be separated; and columns G to L describe the percentage correlation between related reflections. In this example the powder 20 diffraction data has been Pawley fitted using a predetermined unit cell and space group. Reflection intensities have been allowed to vary as variables in a least squares fit to the diffraction data. The result is a file containing estimates of the individual reflection intensities. However, where reflections lie very close together in 20 they may be treated as a clump. 25 Where the reflections are close, but not close enough to be treated as a clump, the correlation terms provide information on how the respective intensities are related. Thus, the first reflection at line 1:

1 0 0 10.047 0.1106 1 0 0 0 0 0 0

is an isolated reflection whereas at lines 18 and 19 two correlated 30 reflections are identified. At lines 41 and 42 two reflections are identified that lie so close together that they are treated as a single variable intensity. With the reduced representation the diffraction data has been effectively

condensed into the data contained in columns D and E in combination with non-zero elements of columns G to L.

The close agreement between the two structures shown in Figure 10 demonstrates one characteristic of the method described above - the 5 ability of the simulated annealing method to 'fine tune' the structure, generally finding a solution very close to the global minimum in χ^2 space. This structure solution, which involved the optimisation of 16 degrees of freedom (10 internal, 6 external) took approximately 40 minutes to execute on a DEC Alphastation 500/500.

10 In another analysis, the molecule under study is said to possess a total energy equal to the sum of its potential energy (proportional to the current χ^2) and some kinetic energy applied to start the molecule moving over the χ^2 hypersurface. The gradient of the χ^2 hypersurface with respect to each of the degrees of freedom is calculated and it is these gradients 15 that control the resultant trajectory of the molecule and thus the way in which it explores the hypersurface. Lowering the temperature of the system in a way analogous to that already described gradually decreases the kinetic energy of the system and that in turn leads to the molecular configuration conforming to the global minimum of the χ^2 hypersurface.

20 In a further adaption of the method, regardless of the global optimisation strategy used, both local and semi-global optimisation methods (e.g. Newton-Raphson, simplex) can be invoked when the χ^2 value reaches some predetermined value that is anticipated to be in the vicinity of the global minimum, thus providing accelerated convergence.

25 With the method and apparatus described above, molecular crystal structures may be solved from powder diffraction data alone. Definition of the molecular fragments in terms of internal co-ordinates means that for a single molecular fragment, problem complexity scales with the number of variable torsion angles rather than with the number of atoms in the 30 fragment. Thus, complex structures can be represented by quite short chromosomes and solved relatively easily. The simple description of

molecular geometry employed, together with the genetic algorithm / simulated annealing analyses and the specified fitness function has thus been shown to be particularly powerful in determining crystal structures from powder diffraction data in a relatively short time frame.

5 To assist in an understanding of the invention, the method has been described with reference to functional, i.e. analyser/processor units. It will of course be apparent that in practice the method is implemented as a program on a computer. Indeed, one of the advantages of this method is that the program can be implemented on a number of different computer
10 architectures, including personal computers and a network of personal computers/workstations acting as a parallel computer.

CLAIMS

1. A method for determining molecular crystal structures from powder diffraction data comprising the steps of: generating a reduced representation of the powder diffraction pattern in dependence on a predetermined unit cell and space group of the molecule under examination in which the total quantity of diffraction data is significantly reduced whilst maintaining the characteristics of the diffraction data that are representative of the crystal structure under examination; determining a set of variables for describing trial molecular structures, derived from predetermined internal coordinates and said space group; assigning values to said variables thereby creating a population of trial structures each defined by a unique set of values for said variables; calculating a fitness for each trial structure with respect to the reduced representation of the powder diffraction pattern; determining whether any one of the calculated fitnesses is less than or equal to a predetermined threshold; where none of the calculated fitnesses is less than or equal to the threshold value, selecting at least one survivor from the population of trial structures, altering the values of the variables of at least one of the survivors in accordance with one or more predetermined rules, calculating the fitnesses of the new trial structures; and repeating the steps of selecting survivors, altering the values of the variables and calculating the fitnesses of the new trial structures until at least one of the calculated fitnesses is less than or equal to the threshold value, and where at least one of the calculated fitnesses is less than or equal to the threshold, outputting at least one trial molecular crystal structure represented by the successful sets of values.

2. A method as claimed in claim 1, wherein the reduced representation includes single values representative of the intensity of each

reflection in the powder diffraction data and one or more factors representative of the extent to which adjacent reflections overlap.

3. The method as claimed in claim 2, wherein the reduced
5 representation consists of a structure factor intensity listing and associated covariance matrix.
4. The method as claimed in either of claims 2 or 3, wherein the total
10 data in the reduced representation is reduced by a factor substantially equal to the number of data points in the original powder diffraction data divided by the number Bragg reflections in the measured data range.
5. The method as claimed in either of claims 3 or 4, wherein the fitness
15 χ^2 of each of the trial structures is determined using the following function:

$$\chi^2 = \sum_h \sum_k \{ (I_h - c|F_h|^2) (V^{-1})_{hk} (I_k - c|F_k|^2) \}$$

20 where:

$I_{h,k}$ = extracted intensity

V_{hk} = covariance matrix

c = a scale factor

$F_{h,k}$ = calculated structure factor from trial structure

- 25 6. The method as claimed in any one of the preceding claims, wherein the set of variables consists of three co-ordinates representative of the location of the molecule within the unit cell and three independent co-ordinates representative of the orientation of the
30 molecule within the unit cell.
7. The method as claimed in claim 6, wherein the set of variables

includes one or more co-ordinates representative of variable torsion angles, bond angles or bond lengths.

8. The method as claimed in any one of the preceding claims including
5 the step of determining the unit cell and space group for the molecule under examination.
9. The method as claimed in any one of the preceding claims including
10 the step of determining the set of internal co-ordinates.
10. The method as claimed in any one of the preceding claims, further including the step of monitoring the number of iterations in which new trial structures are generated and halting the method and outputting the trial crystal structure with the best calculated fitness
15 after completion of a predetermined number of iterations.
11. The method as claimed in any one of the preceding claims, wherein
20 the selection of survivors and the alteration of the values of the variables is based on a simulated annealing procedure.
12. Apparatus for determining molecular crystal structures comprising a structure factor analyser for generating from experimental powder diffraction data for the molecule under examination a reduced representation of the powder diffraction pattern based on a
25 predetermined unit cell and space group in which the total quantity of diffraction data is significantly reduced whilst the characteristics of the diffraction data representative of the crystal structure under examination are maintained; a controller for determining a set of variables for describing trial molecular structures, derived from predetermined internal co-ordinates and said space group; a
30 searching processor for creating a population of trial structures each defined by a unique set of values for said variables said searching

processor including a fitness analyser for calculating a fitness for each trial structure with respect to the reduced representation of the powder diffraction pattern, a thresholding device for determining whether any one of the calculated fitnesses is less than or equal to 5 a predetermined threshold, a survivor selector for selecting at least one survivor from the population of trial structures, a variable adjustment device for altering the values of the variables of at least one of the survivors and output means for outputting the one or more trial molecular crystal structures having calculated fitnesses 10 less than or equal to the threshold value.

13. Apparatus as claimed in claim 12, wherein the reduced representation includes single values representative of the intensity of each reflection in the powder diffraction data and one or more factors representative of the extent to which adjacent reflections 15 overlap.
14. Apparatus as claimed in claim 13, wherein the structure factor analyser generates a reduced representation consists of a structure 20 factor intensity listing and associated covariance matrix.
15. The method as claimed in either of claims 13 or 14, wherein the total data in the reduced representation is reduced by a factor substantially equal to the number of data points in the original 25 powder diffraction data divided by the number Bragg reflections in the measured data range.
16. Apparatus as claimed in either of claims 14 or 15, wherein the fitness analyser determines the fitness χ^2 of each of the trial 30 structures using the following function:

$$\chi^2 = \sum_h \sum_k \{ (I_h - c|F_h|^2) (V^{-1})_{hk} (I_k - c|F_k|^2) \}$$

where:

$I_{h,k}$ = extracted intensity from the structure factor analyser

V_{hk} = covariance matrix from the structure factor analyser

c = a scale factor

5 $F_{h,k}$ = calculated structure factor from trial structure

17. Apparatus as claimed in any one of claims 12 to 16, wherein
controller determines a set of variables consists of three co-
ordinates representative of the location of the molecule within the
10 unit cell and three independent co-ordinates representative of the
orientation of the molecule within the unit.

18. Apparatus as claimed in any one of claims 12 to 17, wherein the
structure factor analyser additionally determines the unit cell and
15 space group for the molecule under examination.

19. Apparatus as claimed in any one of claims 12 to 18 including a co-
ordinate generator for determining the set of internal co-ordinates.

20. Apparatus as claimed in any one of claims 12 to 19, further
including a counter for monitoring the number of iterations of new
trial structures.

Fig 1.

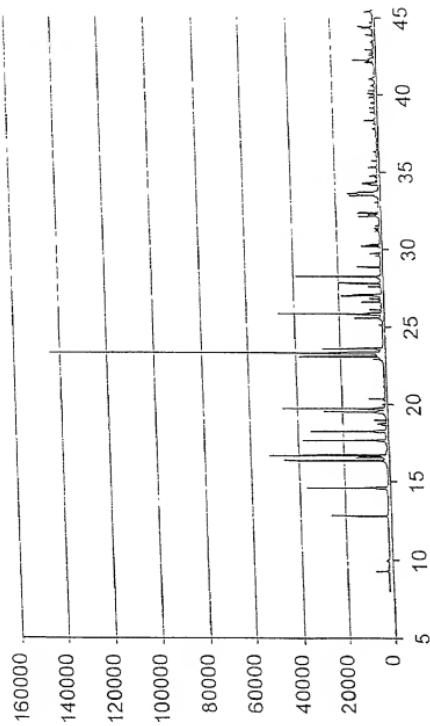
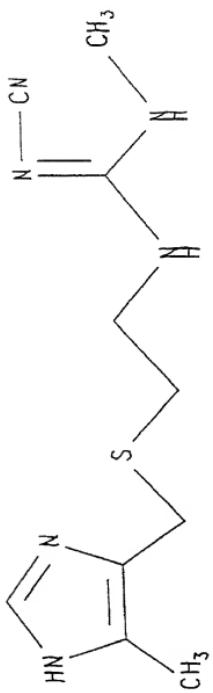


Fig 2.



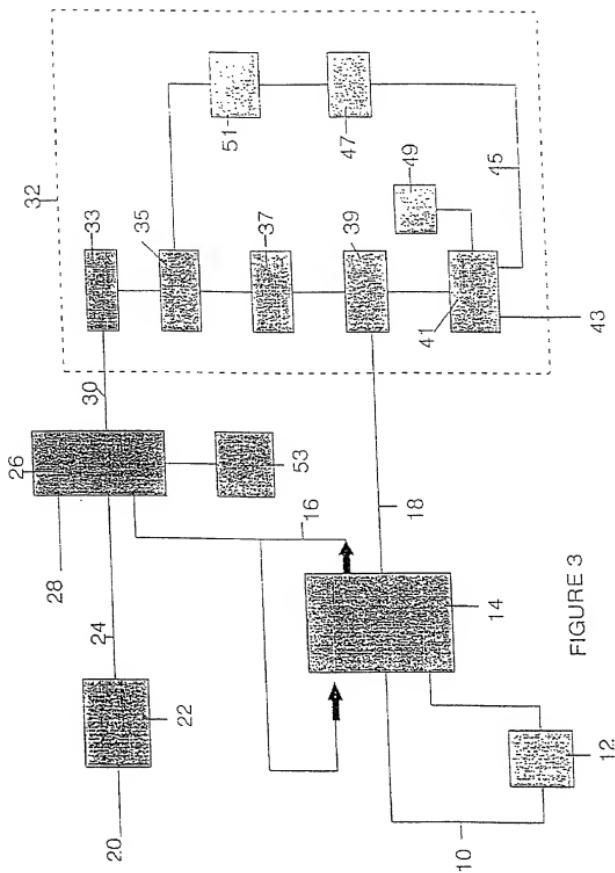


Fig. 4.

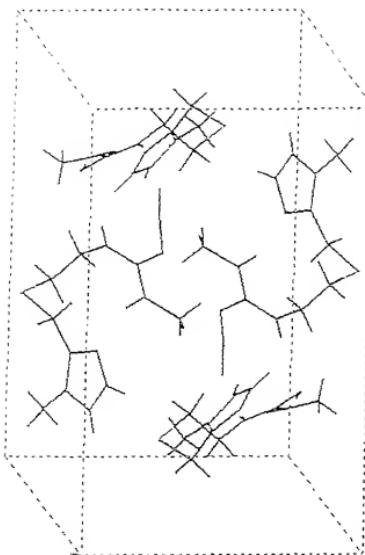
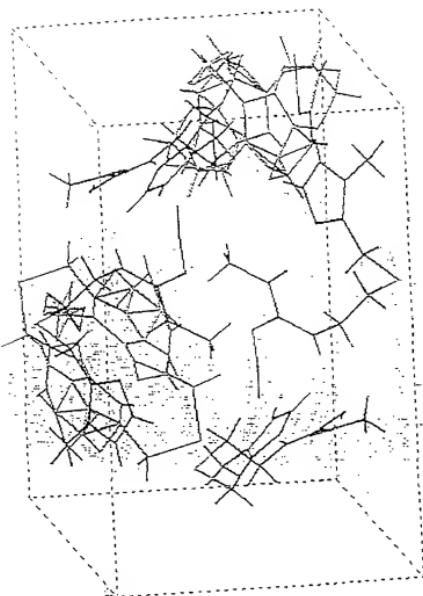


Fig 5a



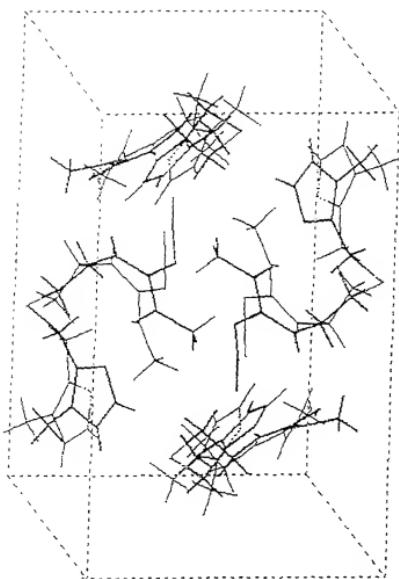
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Fig 5b.



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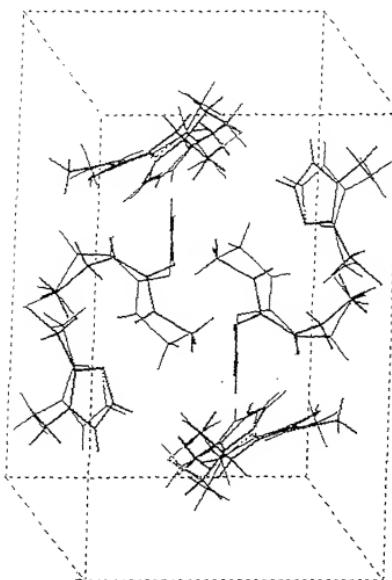
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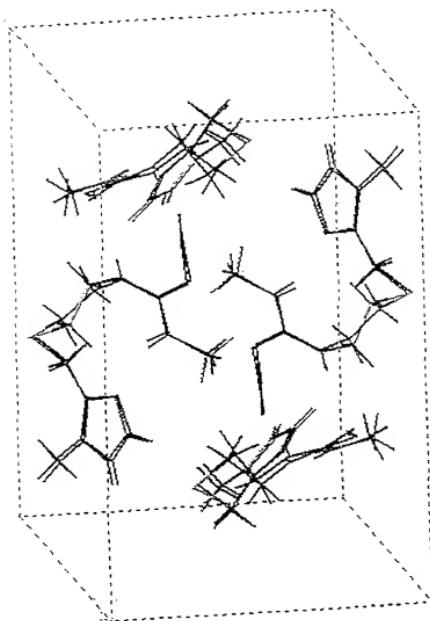
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Fig 5c.



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Fig 5d.



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Fig 6.

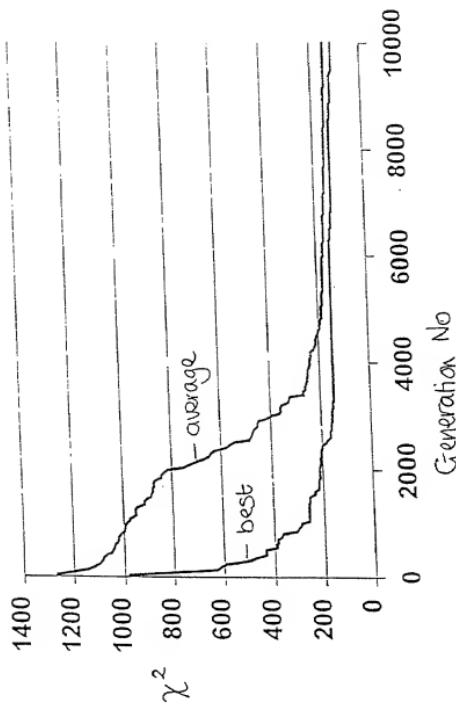


Fig 7a.

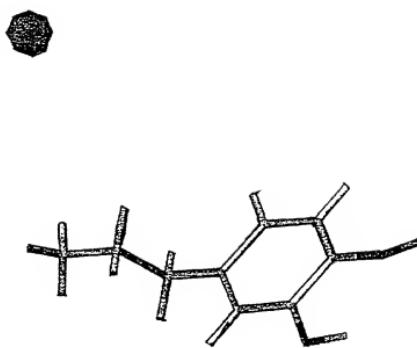


Fig 7b.

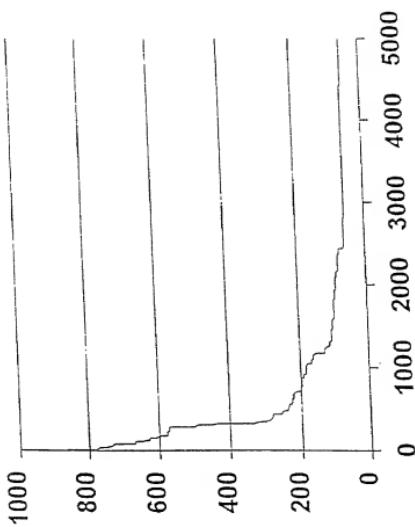
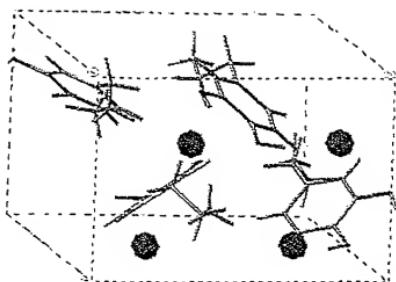
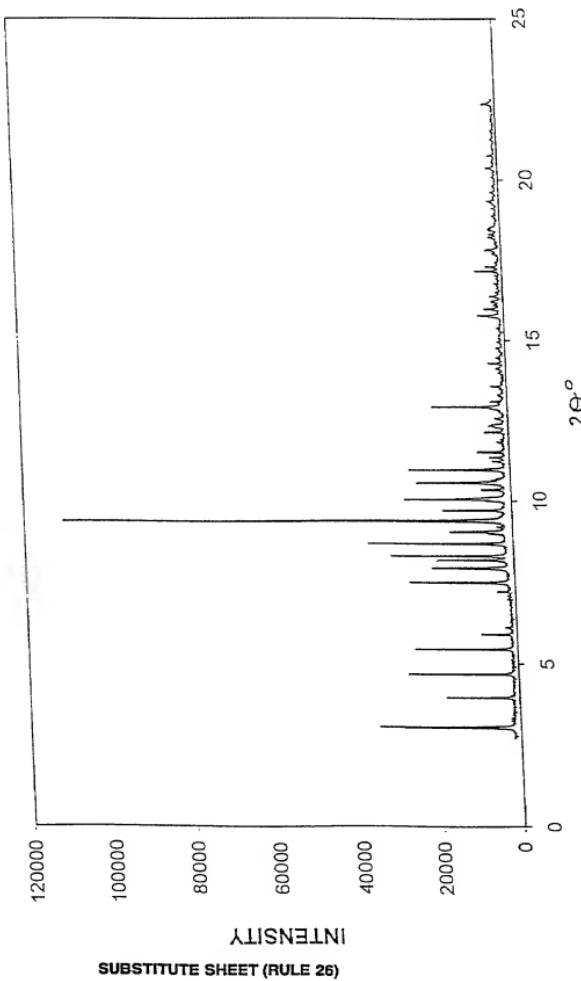


Fig 7c.



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FIGURE 8

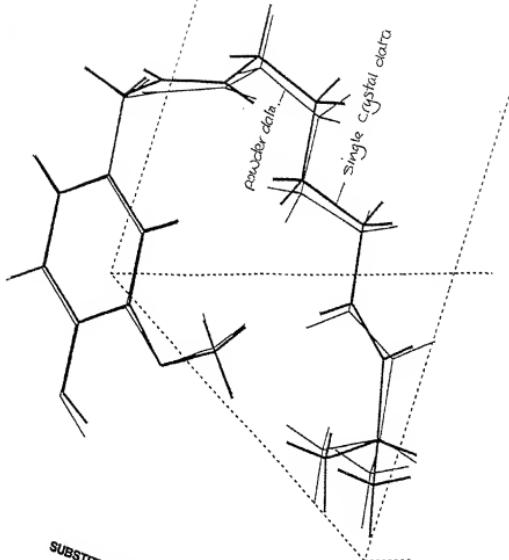


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FIGURE 9

	A	B	C	D	E	F	G	H	I	J	K	L
1	1	0	0	10.047	0.1106	1	0	0	0	0	0	0
2	-1	1	0	3.637	0.0617	2	0	0	0	0	0	0
3	0	1	1	5.764	0.0874	3	0	0	0	0	0	0
4	0	2	0	0.321	0.0666	4	0	0	0	0	0	0
5	-1	1	1	8.983	0.1116	5	0	0	0	0	0	0
6	1	1	1	0.284	0.0432	6	0	0	0	0	0	0
7	-1	2	0	3.956	0.0916	7	0	0	0	0	0	0
8	2	0	0	2.070	0.1117	8	0	0	0	0	0	0
9	0	2	1	-0.025	0.0446	9	0	0	0	0	0	0
10	-1	2	0	0.005	0.0493	10	0	0	0	0	0	0
11	-1	2	1	0.375	0.0660	11	0	0	0	0	0	0
12	1	2	1	2.156	0.0959	12	0	0	0	0	0	0
13	-2	1	1	21.121	0.2266	13	0	0	0	0	0	0
14	0	0	2	10.592	0.4909	14	-55	34	0	0	0	0
15	2	1	1	9.634	0.6372	15	-88	0	0	0	0	0
16	-2	2	0	9.055	0.5427	16	10	0	0	0	0	0
17	-1	3	0	17.781	0.2205	17	0	0	0	0	0	0
18	-1	0	2	8.314	0.5049	18	-57	0	0	0	0	0
19	0	1	2	25.368	0.3353	19	0	0	0	0	0	0
20	0	3	1	0.385	0.0972	20	0	0	0	0	0	0
21	-1	1	2	2.992	0.2855	21	-59	0	0	0	0	0
22	1	0	2	23.776	0.2748	22	0	0	0	0	0	0
23	-2	2	1	23.776	0.2748	22	0	0	0	0	0	0
24	-1	3	1	5.447	0.1809	23	0	0	0	0	0	0
25	1	1	2	10.351	0.3342	24	-59	0	0	0	0	0
26	2	2	1	16.338	0.3505	25	0	0	0	0	0	0
27	1	3	1	0.341	0.1841	26	-69	0	0	0	0	0
28	3	0	0	4.652	0.3947	27	0	0	0	0	0	0
29	0	1	2	137.393	0.6686	28	0	0	0	0	0	0
30	-3	1	0	1.685	0.1391	29	0	0	0	0	0	0
31	-2	0	2	0.293	0.3221	30	-28	0	0	0	0	0
32	-1	2	2	21.709	0.3157	31	-23	0	0	0	0	0
33	-2	3	0	0.647	0.1616	32	0	0	0	0	0	0
34	-2	1	2	6.018	0.2122	33	0	0	0	0	0	0
35	1	2	2	34.450	0.4173	34	-39	0	0	0	0	0
36	-3	1	1	14.637	0.3554	35	-31	0	0	0	0	0
37	0	4	0	12.411	0.4668	36	10	0	0	0	0	0
38	2	0	2	17.335	0.5226	37	-53	0	0	0	0	0
39	-2	3	1	1.605	0.2200	38	10	0	0	0	0	0
40	-3	2	0	2.854	0.1700	39	0	0	0	0	0	0
41	-1	4	0	22.869	0.2070	40	0	0	0	0	0	0
42	3	1	1	22.869	0.2070	40	10	0	0	0	0	0
43	1	2	1	4.788	0.2129	41	-27	0	0	0	0	0
44	2	3	1	1.945	0.1824	42	0	0	0	0	0	0
45	0	4	1	-0.298	0.1358	43	0	0	0	0	0	0
46	-2	2	2	0.088	0.1723	44	0	0	0	0	0	0
47	0	3	2	44.292	0.5083	45	-48	0	0	0	0	0
48	-3	1	2	3.714	0.3270	46	0	0	0	0	0	0
49	-1	4	1	1.319	0.1982	47	-44	0	0	0	0	0
50	-1	3	2	4.787	0.2325	48	0	0	0	0	0	0
51	1	4	1	7.086	0.2216	49	0	0	0	0	0	0
52	3	2	1	-0.509	0.1419	50	0	0	0	0	0	0

Figure 10



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DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are stated below next to my name.

I believe I am the original, first, and sole inventor (if only one name is listed below) or an original, first, and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

TITLE OF INVENTION**METHOD AND APPARATUS FOR DETERMINING MOLECULAR CRYSTAL STRUCTURES**

the specification of which has an international filing date of 31 July 1998 as PCT Application No. PCT/GB98/02316

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with 37 CFR §1.56.

I hereby claim foreign priority benefits under 35 U.S.C. §119(a)-(d) or §365(b) of any foreign application(s) for patent or inventor's certificate, or §365(a) of any PCT international application which designated at least one country other than the United States, listed below and have also identified below any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN/PCT APPLICATION(S)

COUNTRY/OFFICE	APPLICATION NO.	DATE OF FILING	PRIORITY CLAIMED
Great Britain	9716278.8	31 July 1997	<input checked="" type="checkbox"/> YES NO <input type="checkbox"/>
			<input type="checkbox"/> YES NO <input type="checkbox"/>
			<input type="checkbox"/> YES NO <input type="checkbox"/>
			<input type="checkbox"/> YES NO <input type="checkbox"/>

I hereby claim the benefit under 35 U.S.C. §119(e) of any United States provisional application(s) listed below.

PROVISIONAL APPLICATION NUMBER

DATE OF FILING

None

None

I hereby claim the benefit under 35 U.S.C. §120 of any United States application(s) or §365(c) of any PCT International application(s) designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of 35 U.S.C. §112, I acknowledge the duty to disclose material information as defined in 37 CFR §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

**PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS
DESIGNATING THE U.S. FOR BENEFIT UNDER 35 U.S.C. §120**

Application Serial No.	Date of Filing	Patented	Pending	Abandoned	Status (check one)
none		<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	
		<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	

And I hereby appoint Arthur H. Seidel, Registration No. 15,979; Gregory J. Lavorgna, Registration No. 30,469; Daniel A. Monaco, Registration No. 30,480; Thomas J. Durling, Registration No. 31,349; and John J. Marshall, Registration No. 29,671, my attorneys or agents with full power of substitution and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

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